

Specificity of Coproporphyrinogen Oxidase: Conversion of Coproporphyrinogen-IV into Protoporphyrin-XIII

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Summary Coproporphyrinogen-IV (**1b**) is converted into protoporphyrin-XIII by avian haemolysates, and the structure of the product confirmed by total synthesis.

WE have shown¹ how coproporphyrinogen-III (**1a**) is converted by avian haemolysates and rat liver homogenates by a specific pathway into protoporphyrin-IX. Coproporphyrinogen-IV (**1b**) is also a substrate for the enzyme involved, coproporphyrinogen oxidase [EC1.3.3.3], whereas coproporphyrinogens-I and -II are not.²

There is indirect evidence³ that pentacarboxylate porphyrinogen-III (**1c**) is also a substrate for this enzyme, whereas the product of this reaction, dehydroisocoproporphyrinogen (**2c**) is not. Comparison of the structures of

these six compounds suggests that the minimum structural requirement necessary for a substrate is the sequence [methyl† methyl-propionate methyl]. We therefore reasoned that the product from coproporphyrinogen-IV would be protoporphyrin-XIII (**2b**).[‡]

Coproporphyrinogen-IV (*ca.* 2 mg) was shaken with a haemolysate⁵ of chicken erythrocytes at 37 °C for 3 h in the dark and the product (*ca.* 1 mg) after isolation in the usual way was converted into the methyl ester. T.l.c. and h.p.l.c.⁶ showed that it was essentially a single compound with two ester side-chains. Field desorption and electron impact mass spectrometry gave a molecular ion (*m/e* 590) corresponding to a protoporphyrin dimethyl ester, and n.m.r. spectroscopy in the presence of the shift reagent

† Can be replaced by H or vinyl.

‡ Professor B. Frydman (Buenos Aires) independently reached the same conclusions.

[Eu(fod)₃] showed that the two remaining ester groups flanked one *meso*-position (as the corresponding *meso*-proton experienced a very marked downfield shift⁷). Of the

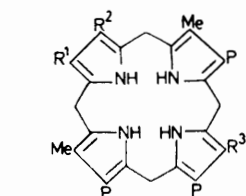
decarboxylation of two propionate groups) only one, the XIII isomer (3), retains this arrangement of propionate ester groups.

Protoporphyrin-XIII dimethyl ester (3) was synthesised from the two pyrromethanes (4) and (5) followed by conversion⁸ of the hydroxyethyl side-chains into vinyl groups. The product (3), m.p. 198–200 °C, was identical with that obtained from coproporphyrinogen-IV by mixed m.p. and 'mixed' n.m.r. spectra in the presence of the europium shift reagent;⁹ e.g. the *meso* protons gave rise to a 1:2:1 triplet at τ ca. -0.2 to -0.4 (a 3:1 doublet at low concentration) and on addition of shift reagent (5 mol. equiv.) a singlet moved downfield to τ ca. -6 whereas the other signals were relatively little affected.

Studies with the enzyme present in liver homogenates show that an intermediate tricarboxylic porphyrinogen is formed in appreciable amounts (ca. 35%) *en route* to protoporphyrin-XIII, and this is confirmed by kinetic experiments with haemolysates.¹⁰ The amount of hardoporphyrin which can be isolated from haemolysate experiments with coproporphyrinogen-III is <10% of added substrate.¹¹ In contrast to previous findings,² however, both coproporphyrinogens-III and -IV are metabolised at similar overall rates to the corresponding protoporphyrins. Full details of these and related experiments will be published elsewhere; the synthesis of the tricarboxylic porphyrin derived from coproporphyrinogen-IV is in progress.

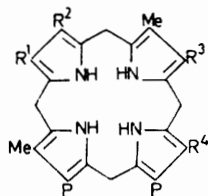
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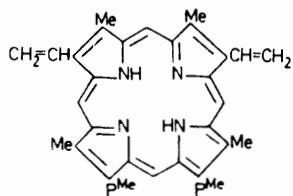
(1)

- a: $R^1 = R^3 = \text{Me}$, $R^2 = \text{P}$
 b: $R^1 = \text{P}$, $R^2 = R^3 = \text{Me}$
 c: $R^1 = \text{Me}$, $R^2 = \text{P}$, $R^3 = \text{CH}_2\text{CO}_2\text{H}$

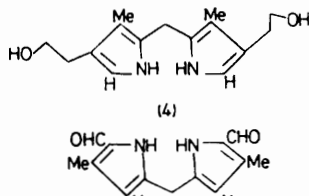
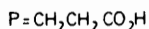


(2)

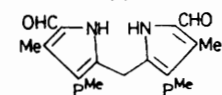
- a: $R^1 = R^4 = \text{Me}$, $R^2 = R^3 = \text{CH}=\text{CH}_2$
 b: $R^2 = R^4 = \text{Me}$, $R^1 = R^3 = \text{CH}=\text{CH}_2$
 c: $R^1 = \text{Me}$, $R^2 = \text{CH}=\text{CH}_2$, $R^3 = \text{P}$,
 $R^4 = \text{CH}_2\text{CO}_2\text{H}$



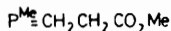
(3)



(4)



(5)



four possible protoporphyrin isomers which might have been formed from coproporphyrinogen-IV (by oxidative

¹ D. E. Games, A. H. Jackson, J. R. Jackson, R. V. Belcher, and S. G. Smith, preceding communication.

² R. J. Porra and J. E. Falk, *Biochem. J.*, 1964, **90**, 69; S. Granick and R. D. Levere, *Progr. Haematol.*, 1964, **4**, 1.

³ G. H. Elder, *Biochem. J.*, 1972, **126**, 877.

⁴ G. H. Elder and J. O. Evans, unpublished results.

⁵ Cf. E. I. B. Dresel and J. E. Falk, *Biochem. J.*, 1956, **63**, 388.

⁶ N. Evans, D. E. Games, A. H. Jackson, and S. A. Matlin, *J. Chromatog.*, 1975, **115**, 325.

⁷ M. S. Stoll, G. H. Elder, D. E. Games, P. O'Hanlon, D. S. Millington, and A. H. Jackson, *Biochem. J.*, 1973, **131**, 429.

⁸ Cf. R. P. Carr, A. H. Jackson, G. W. Kenner, and G. S. Sach, *J. Chem. Soc. (C)*, 1971, 487.

⁹ A. H. Jackson, H. A. Sancovich, A. M. Ferramola, N. Evans, D. E. Games, S. A. Matlin, G. H. Elder, and S. G. Smith, *Phil. Trans. B.*, 1976, **273**, 191.

¹⁰ Cf. R. B. Frydman and B. Frydman, *FEBS Letters*, 1975, **52**, 317.

¹¹ This result confirms earlier findings: cf. S. Sano and S. Granick, *J. Biol. Chem.*, 1961, **236**, 1173, and ref. 2.